

A Secondary Structure Prediction of the Hemorrhagic Metalloprotease Family

Dietlind L. Gerloff, Thomas F. Jenny, Lukas J. Knecht, and Steven A. Benner

Laboratory for Organic Chemistry and Institute for Scientific Computation
E.T.H. Zurich, CH-8092 Switzerland

Received June 7, 1993

Summary: A secondary structure has been predicted for the hemorrhagic metalloproteases using a method developed in Zurich that extracts structural information from patterns of conservation and variation in homologous protein sequences. This prediction tests the limits of the method when applied to a small number of homologous sequences that have undergone only modest evolutionary divergence. Predictions were also obtained using a neural network developed by Sander and coworkers, to date the best fully automated method for predicting secondary structure, and using the classical Chou-Fasman and GOR heuristics. The predictions are different. No crystal structure is known within this protein family, but one is expected shortly. Therefore, this prediction should contribute significantly to the evaluation of the relative merits of these prediction methods. © 1993 Academic Press, Inc.

A method developed at the E.T.H. in Zurich has recently been used to make *bona fide* predictions of various aspects of the conformation of a number of families of proteins [1,2,3,4,5]. This approach has been controversial. To some, the predictions have been "remarkably accurate" [6], finding "core secondary structures much better than ... achieved with standard methods" and providing a "possibility of extending prediction to a tertiary fold [that] is much better" than the per residue score might imply [7], and represent "a spectacular achievement ... that will come to be recognized as a major breakthrough." [8] Others have regarded these comments as "exaggerated" [9], cautioned that "one swallow does not make a summer" [10], questioned the value of *bona fide* predictions in any case, and suggested that because all prediction work is fundamentally statistical, any method that approaches proteins as individual cases violates important rules of procedure in the structure prediction field [10]

Elsewhere, we have made the case that the most productive approach to understanding conformation in proteins is likely to follow procedures that organic chemists have used to address conformational issues in organic molecules generally [1-5, 11]. We recognize that this case will not be universally convincing. Therefore, we present here the first paper where secondary structure predictions by competing methods (the E.T.H. method, a neural network method [12,13], and classical procedures such as Chou-Fasman [14] and GOR [15]) are made for a family of proteins where no crystal structure is available, but where one is imminent.

For this test, we have chosen the hemorrhagic metalloproteases from snake venom [16], kindly brought to our attention by Prof. Edgar Meyer (Texas A & M University). A crystal structure of a representative of this protein is imminent. Further, the protein family contains

only 7 sequences in two subfamilies, with a maximum evolutionary distance of only 78 accepted point mutations per 100 amino acids (PAM units)[17]. Therefore, this protein family appears ideal for testing the scope of various methods that build consensus prediction with protein families having only few sequences and relatively little evolutionary divergence. This contrasts with models for families used to predict structures for protein kinase [1], the SH3 domain [2], the MoFe nitrogenase protein [3], and others [4], where at least 10 and often over 50 sequences were used as input, with rather large sequence divergence in several cases (e.g., for the MoFe nitrogenase, the tree was 174 PAM units wide).

MATERIALS AND METHODS

The E.T.H. prediction was made by assigning surface, interior, active site, and parsing residues in the alignment [1-4]. The first three assignments convey tertiary structural information. An unrefined consensus secondary structure is assigned to the alignment based on an analysis of patterns in these assignments, together with special effort to accommodate secondary structures that do not readily reveal themselves by such methods (e.g., internal alpha helices) [1]. Predictions using the neural network developed by Sander and coworkers [12,13] were obtained through a server maintained by these workers in Heidelberg. A consensus prediction for the protein family was kindly provided by B. Rost and C. Sander. Chou-Fasman [14] and GOR [15] predictions were obtained routinely. A consensus prediction was obtained in each following a procedure similar to that introduced by Kirschner *et al.* [18] in their prediction of the tryptophan synthase.

RESULTS AND DISCUSSION

The predictions are shown in Figure 1, with the consensus predictions derived in each method shown in bold. Consensus predictions assume that homologous proteins have similar secondary structures [19]. Given the relatively small overall sequence divergence in this family, this assumption is highly plausible. However a consensus prediction does not correspond exactly to any individual member of the family (although the structure of each member might in principle be predicted from a consensus prediction by homology modelling).

Predictions made by the different methods are different, often markedly so. Indeed, all four methods predict the same secondary structure at only 37% of the residues. Thus, an experimental structure should not only permit evaluation of the relative merits of the prediction methods (at least in this test case), but also indicate whether more reliable predictions might be obtained from a composite prediction method involving all four methods.

Regarding supersecondary and tertiary structure, the small number of sequences in the multiple alignment makes it difficult to assemble a complete model for the protein family from the secondary structure prediction made by the E.T.H. method, primarily because sequence pairs at an evolutionary distance appropriate for performing covariation analysis are not available [1]. Nevertheless, a surface antiparallel beta sheet including strands in positions 50-68 is strongly indicated, flanked by at least one additional strand in position ca. 95-104. Also indicated is an active site flanked by three helices, one containing putative zinc binding ligands (positions 133-145) and two containing putative substrate binding units (positions 33-48, and positions 175-193). Interestingly, the active site helix, assigned using rules developed following the incorrect assignment of an internal helix in protein kinase [1], is suggested by analogy with thermolysin [20] and astacin [21], two well studied metalloproteases dependent on

No.	Sequences			ETH Prediction					Neural Net				GOR				Chou-Fasman			
	abcd	efg		1	2	3	4	5	abcd	efg	C	abcd	efg	C	abcd	efg	C			
001	QQ-Q	-Q-	?																	
002	RRRR	NN-	fil spl	A																
003	FFFF	LL-	inside	A																
004	PPPP	PP-	P				p													
005	QQQR	QQQ	surf	A																
006	RRRR	SRK	surf	A																
007	YYYY	YYY	inside	A					BBBB	BBB	B									
008	IIII	III	inside	A	b			b	BBBB	BBB	B									
009	KEEK	EEE	surf	A	b			b	BBBB	BBB	B									
010	LLLL	LLL	inside	A	B			B	BBBB	BBB	B									
011	GAAA	VVV	inside	A	B			B	BBBB	BBB	B									
012	IIII	VVV	inside	A	B			B	BBBB	BBB	B									
013	FVVV	VVV	inside		B			B	BBBB	BBB	B									
014	VVVV	AAA	inside		B			B	BBBB	BBB	B									
015	DDDD	DDD	D					\$	BBBB	BBB	B									
016	HHHH	HHH	H					\$												
017	GGGG	RRG	surf	A				A		BBB										
018	MMMI	MVM	INSIDE	A				A	BBBB	BBB	B									
019	YYVY	FFF	inside	A				A	BBBB	BBB	B									
020	TTKT	MMT	amphi	A	s			A	BBBB	BBB	B									
021	KKKK	KKK	K	A	s	b		\$	BBBB	BBB	B									
022	YYHY	YYY	inside	A	b			A	BBBB	BBB	B									
023	SSNH	NNN	surface	A	b	p		A												
024	GSQG	SSG	Hb var	A	b	p		A												
025	NNNN	DDN	surface	A	b	p		A												
026	SFSL	LLL	inside	A	b	p		A												
027	EKDK	NNN	surface	A	b			A	A A	AAA	A									
028	RKKK	TTT	surface	A				A	AAAA	AAA	A									
029	IIII	III	inside	A				A	AAAA	AAA	A									
030	TRKR	RRR	surface	A				A	AAAA	AAA	A									
031	KKVK	TTT	surface	A				A	AAAA	AAA	A									
032	RRRW	RRR	surface	AA					AAAA	AAA	A									
033	VVVI	VVV	inside	AA				A	BBAA	AAA	A									
034	HHHY	HHH	?	A-				A	BBAA	AAA	A									
035	QQQQ	EEE	fil spl	A				A	BBAA	AAA	A									
036	MMML	III	inside	A				A	AAAA	AAA	A									
037	IVVV	VVV	inside	A				A	AAAA	AAA	A									
038	NNNN	NNN	N	A				\$	A	AAAA	AAA	A								
039	NNHT	FFT	amphi	A	p			A	A	AAA	A									
040	IIII	IIL	inside	A				A	AAAA	AAA	A									
041	NNNN	NNN	N	A				\$	A	AAAA	AAA	A								
042	MEEN	EGG	SURFACE	A	p			A	AAAA	AAA	A									
043	MMMI	FFF	inside	A				A	AAAA	AAA	A									
044	CYYY	YYY	inside	AA				A	AA	AA	A									
045	RRRR	RRR	R	AA				\$	A											
046	APPS	SSS	Hb var	AA																
047	LLLL	LLL	inside	AA					BBBB	BBB	B									
048	NNNN	NNN	N	AA				\$												
049	IIII	III	inside	A	B			b	BBBB	BBB	B									
050	VAAL	RHL	surface	A	B			b	BBBB	BBB	B									
051	TIIV	VVI	INSIDE	A	B			b	BBBB	BBB	B									
052	TTSA	SSS	Hb var	A	B			b	BBBB	BBB	B									
053	LLLL	LLL	inside	A	B			b	BBBB	BBB	B									
054	SSNV	TTT	surface	A	B			b												
055	VLRY	DDD	surface		B			B	BBBB	BBB	B									
056	LLLL	LLL	inside		B			B	BBBB	BBB	B									
057	EDQE	EEE	surface		B			B	BBBB	BBB	B									
058	IVII	III	inside		B			B	BBBB	BBB	B									
059	WWWW	WWW	inside		b				BBBB	BBB	B									
060	SSSS	SSS	S					\$	BBBB	BBB	B									

Figure 1. A multiple alignment for seven hemorrhagic metalloproteases (alignment position numbers and sequences shown vertically, one letter code used, underlining indicates parsing string); Accession numbers (SwissProt): (a) P15503; (b) P14530; (c) P20165; (d) P20164; (e) P20897; (f) P15167; (g) P22796, followed by secondary structure predictions. "A", "a", "B", "b", "T" and "t" indicate strong and weak alpha helix, beta strand, and turn assignments, respectively. A "?" indicates that an assignment is uncertain. E.T.H. prediction [1-5]. Column 1 shows surface/interior assignments made using the E.T.H. method (underlining indicates consecutive surface assignments indicative of breaks in secondary structure); "fil spl": hydrophilic split; "amphi": amphiphilic position separating two branches in tree; "Hb var": hydrogen bonding variable. Column 2 shows possible alpha helix assignments made using the

061	EEKK	DNN	SURFACE						p	AATT	T.T	T	AATa	TtT	T	
062	KKKQ	QEQ	SURFACE							AATT	T.T	T	AATa	TTT	T	
063	DDDN	DDD	surface							AATT	TTT	T	AABB	TTT		
064	LLLK	FQL	amphi		B	B		B		AABT	BT.		BABB	BBB	B	
065	IIII	III	inside		B	B	BBBB	BBB	B	AABB	BT.		BABB	BBB	B	
066	TTTT	TNN	surface		B	B	BBBB	BBB	B	AABB	B..		BABB	BBB	B	
067	VMVV	VIV	INSIDE		B	B	BBBB	BBB	B	AABB	B..		BABB	BBB	B	
068	QKQK	QQQ	surface		B	B	BBBB	BBB	B	AABB	...		BA.B	BBB	B	
069	AASS	SSS	Hb var		B	p	B	BBBB	BBB	B	.A.B	...	BA..	...		
070	SVAA	SAA	Hb var	a	b	p			Att	.t.		
071	AASS	ASA	Hb var	a	b	p			Att	.t.		
072	PPNN	KSN	SURFACE	A	b	p	A	A	A	At.B	tT.		
073	TTVV	NDD	surface	A		p	A	AAAA	AAA	A	.BBB	BtaB	tT.		
074	TTTT	TTT	T	A		\$	A	AAAA	AAA	A	BBBB	...B	BbaB	bbB	B	
075	LALL	LLL	inside	A	B		A	AAAA	AAA	A	BBBB	...B	BbaB	bbB	B	
076	TRED	HNK	SURFACE	A	B		A	AAAA	AAA	A	BBBB	...B	BbaB	bbB	B	
077	LLSL	SAT	Hb var	A	B		A	AAAA	AAA	A	BB.B	.A.	BbaB	bbB	B	
078	FFFF	FFF	inside	A	B		A	AAAA	AAA	AA..	BbaB	bbB	B	
079	GGGG	GAG	inside	A	B	p	A	AAA	AA	AA..	.tTt	.Aa		
080	ADND	EEE	surface	A	B	p	A	AAAA	AAA	A	AAA	AtTt	aAa	A	
081	WWWW	WWW	inside	A	B		A	AAAA	AAA	A	...A	AAA	Aaaa	aAa	A	
082	RRRR	RRR	R	A		\$	A	AAAA	AAA	A	.A.A	AAA	Aaaa	aAa	A	
083	EEEE	KEE	surface	A			A	AAAA	AAA	A	.A.A	AAA	Aaaa	aAa	A	
084	TTTS	STR	SURFACE	A			A	AAAA	AAA	A	.A.A	AAA	Aaaa	aAa	A	
085	VVVV	VDV	surface	a	i	b		AAAA	AAA	A	.A.A	AAA	Aaaa	aAB	A	
086	LLLL	LLL	inside	a	b			AAAA	AAA	A	.A.A	AAA	Aaaa	aAB	A	
087	LLLL	LLL	inside		b			AAAA	AAA	A	.ATA	AA.	Aaaa	aAB	A	
088	NKKK	NNN	surface		b	p		AAA	AAA		TATA	TAT	T	.aaa	.B	
089	RQQQ	RRR	surface			p					TATT	TTT	T	taaa	ttB	
090	TKQR	KKI	SURFACE			p					TATT	TTT	T	ttta	ttB	T
091	SDNS	RSS	SURFACE			p					.ATT	T..		ttta	...	
092	HHNH	HHH	surface			p					.ATT	T..		taTt	.tt	T
093	DDDD	DDD	D		b	\$.ATT	T..		.aTt	TTT	T
094	HHCC	NNN	amphi		b	p		AA	AA	A	.ATT	.A.		.aTt	TTT	T
095	AAAA	AAA	inside	n	B		B	AAAA	AAA	A	.ABT	.A.	BaaB	BAA	A	
096	QQHQ	QQQ	??		o	B	B	AAAA	AAA	A	BABB	BAB	B	BaaB	BAA	A
097	LLLL	LLL	inside		B		B	AAAA	AAA	A	BABB	BAB	B	BaaB	BAA	A
098	LLLL	LLL	inside	3	B		B	BBBB	BBB	B	BBBB	BAB	B	BaaB	BAA	A
099	TTTT	TTT	T		B	\$	B	BBBB	BBB	B	BBBB	BAB	B	BaaB	BAA	A
100	ADAT	AAA	surface	6	B		B	BBBB	BBB	B	BBBB	BAB	B	BaaB	BAA	A
101	TITI	III	inside		B		B	BBBB	BBB	B	BB.B	BAB	B	BaaB	BAA	A
102	INND	VED	SURFACE		B		B	BBBB	BBB	B	B..T	BAB		B..B	BAA	
103	FFLF	LLL	inside		B		B				B..T	BAB		B..B	BAA	
104	NTND	DDA	SURFACE	A	B	p	B				T..T	BA.		tttt	TAA	T
105	GGDG	DED	SURFACE	A		p					T... BA.			ttTt	TAt	T
106	NNNP	YEN	SURFACE	A		p				 BA.			ttTt	tAt	T
107	VTTT	TTT	inside	A	B		?			 BA.			.BBt	bbB	B
108	IIII	LLI	inside	A	B		?	BBBB	BBB	B BAB			.BB.	bbB	B
109	GGGG	GGG	G	A	B	p	?	BBBB	BBB	B	T... BAB			.BB.	bbB	B
110	RWLK	LLI	amphi	A	B		?	BBBB	BBB	B	T..B	BAB		.BB.	bbB	B
111	AAAA	AAA	inside	A	B		?	BBBB	BBB	B	.TB	B.B		.BB.	bbB	B
112	PYYY	YPY	inside	A	B	p	?	BBBB	BBB	B	TTTB	T.B	T	tBBb	btB	B
113	VMKT	LLT	SURFACE	A				BBBB	BBB	B	TTTB	TTT	T	tBtb	btB	B
114	GGKA	NGG	SURFACE	a		p					TTTB	TTT	T	t.tb	b.t	
115	GGGS	STG	Hb var	a		p					TTTB	TBT	T	...b	.t	T
116	MMMM	MMM	inside					BB	B		TTTB	.BT	T	tttb	.t	T
117	CCCC	CCC	C			\$.TTB	.BT		tttb	.t	T
118	DNND	HDY	SURFACE			p				B.		
119	PAPP	PPP	inside			p					TTTT	TTT	T	TtTT	TTT	T
120	KKKK	RKK	surface								TTTT	TTT	T	TtTT	TTT	T
121	RNLR	NLN	SURFACE		B	p					T..T	T.T	T	ttBt	T.T	T

E.T.H. method. Trailing character "s" indicates that amphiphilicity is preserved only if the position is assigned to the surface, while trailing character "i" indicates that amphiphilicity is preserved only if the position is assigned to the inside. Unassigned is a coil. Column 3 shows possible beta strand assignments made using the E.T.H. method. Column 4 shows parses made using the E.T.H. method ("p" indicates a strong parse, "p" a weak parse) and active site assignments (\$). Column 5 shows consensus secondary structure assignments made using the E.T.H. method. **Neural net prediction.** [12,13]. Provided by Rost and Sander. Consensus prediction under **C. GOR Prediction.** According to reference [15]. Consensus prediction under **C. Chou-Fasman Prediction.** According to reference [14]. Consensus prediction under **C.**

No.	Sequences abcd efg	ETH Prediction					Neural Net				GOR				Chou-Fasman			
		1	2	3	4	5	abcd	efg	C	abcd	efg	C	abcd	efg	C			
122	SSSS SSS S		B	\$	B					B.BT	T.T		.tB.	...				
123	VVVV VIV inside		B		B	B	BBBB	BBB B		BBBB	TBB B		BtBB	BBB B				
124	AGGG GGG inside		B		B	B	BBBB	BBB B		BBBB	BBB B		B.BB	BBB B				
125	IILI LII INSIDE		B		B	B	BBBB	BBB B		BBBB	BBB B		B.BB	BBB B				
126	VVVV IVV inside		B		B	B	BBBB	BBB B		BBBB	BBB B		B.BB	BBB B				
127	RKQQ QQQ surface		B		B	B	BBBB	BBB B		BBBB	BBB B		B.BB	BBB B				
128	DDDD DDD D				\$			B B		...Btt	ttt T				
129	HHYY HHH inside										tttt	ttt T				
130	NSSS SSS surface				p						tTtt	... T				
131	ASPP PPP Hb var				p				T		BTT.	..T				
132	INNI IIK SURFACE				p			B B		B...	..T		BTB	BBT B				
133	VVVN NNT SURFACE A					A	BBBB	BBB B		BAAB	..A		BAAB	BBB B				
134	FFFL LLL inside A					A	BBBB	BBB B		BAAB	..A		BAAB	BBB B				
135	VMMV LLL inside A					A	BBBB	BBB B		BAAB	AAA A		BAAB	BBB B				
136	VVVV MMI inside A					A	BBBB	BBB B		AAAB	AAA A		BAAB	BBB B				
137	AAAA GGA inside A					A	BBBB	BBB B		AAAA	AAA A		BAAB	aaB A				
138	VVVV VVV inside A					A	BBBB	BBB B		AAAA	AAA A		BAAB	aaB A				
139	TTTI TTT inside A					A	BBBB	BBB B		AAAA	AAA A		BAAB	aaA A				
140	MMMM MMM inside A					A	BBBB	BBB B		AAAA	AAA A		BAAB	aaA A				
141	TTTT AAA inside A					A	BBBB	BBB B		AAAA	AAA A		.AA.	aaA A				
142	HHHH HHH H					A	AAAA	AAA A		AAAA	AAA A		.AA.	aaA A				
143	EEEE EEE E					\$	A	AAAA	AAA A	AAAA	AAA A		.AA.	aaA A				
144	MILM LLL inside A					A	AAAA	AAA A		AAAA	AAA A		.AA.	aaA A				
145	GGGG GGG G				A					AAAT	AAA A					
146	HHHH HHH H					\$				A..T	AAA A					
147	NNNN NNN N					\$				A...	..A					
148	LLLL LLL inside					B		B		A..	..A		aaa .				
149	GGGG GGG G					B	p	B		AAA.	..A A		aaa .				
150	MMMI MMM inside					B		B		AAA.	TTA A		AAA.	aaa A				
151	HEEP EEK SURFACE					B		B		AAA.	TTA A		AAAt	aaa A				
152	HHHH HHH H					B	\$	B		AAAT	TTA A		AAAt	aaa A				
153	DDDD DDD D					\$				AAAT	TTA A		AAAT	aaa A				
154	EDDG GGE SURFACE					p				AAAT	TTA A		AAAT	..a A				
155	-KK- KKN surface					p				-AA-	TTA -		-AA-	TT. -				
156	DDDN DDH SURFACE					p				TAAT	TTT T		A..T	TTT -				
157	KKKS --- surface					p				TAAT	---		TTTT	---				
158	CCCC CCC C					\$				TAAT	TTT T		TTTT	..t T				
159	NKKT LLH SURFACE									TAAT	TTT T		tttT	..t T				
160	CCCC RRC surface									TAAT	TTT T		tttT	..t T				
161	NEEQ GGS SURFACE					p				TAAT	TTT T		B..T	...				
162	TAAG AAA inside					p				TAAT	TTT T		BBBT	..b B				
163	---F SSS ?				a		p			---	..T -		---B	..b -				
164	---P LLF inside				a		p			---	T BBT -		---B	BBb -				
165	CCCC CCC C				a	B	\$	B		BAAT	BBT		BBBB	BBb B				
166	IIII III inside				a	B		B		BAAB	BBT B		BBBB	BBb B				
167	MMMM MMM inside				a	B		B		BAAB	BB. B		BBBB	BBb B				
168	SSSS RRP surface				a	B	p	B		BAAB	BB. B		.BB.	BB. B				
169	KADP PPP surface				a		p		BB	BAAB	TT.		aBB.	TTT				
170	VVVM GGS Hb var				a	B	p	B		BAAT	TT.		aBB.	TTT				
171	LIII LLI INSIDE				a	B		B		BAAT	...		aBB.	...				
172	SSSS TTS Hb var				a	B		B		.TT.	...		att.	...				
173	RDDD RKE SURFACE				a	B		B		.TT.	...		att.	TtT T				
174	QKKE GGG surface					p					a..T	TtT T				
175	PPPE RRP SURFACE				A		p	A		TTT.	...		TTTT	TTT T				
176	SSSS SSS S				A		\$	A		TTT.	..T T		TTTT	TTT T				
177	KKKE YYY surface				A			A		TTTT	..T T		t...t	..t .				
178	YLLL EEE amph				A			A	BBBB	BBB B		TTTT	..T T				
179	FFFF FFF inside				A			A	BBBB	BBB B		TTTT	..T T				
180	SSSS SSS S				A		\$	A		TTTT	..T T		.TTT	..t T				
181	EDDN DDD surface				A		p	A		TTTT	..T T		tTTT	..T T				
182	CCCC ADC surface				A			A		TTTT	..T T		tTTt	..T T				

Figure 1 - Continued

zinc. Despite this similarity, we have been unable to present a convincing case based on either sequence alignment or a secondary structure prediction that hemorrhagic metalloproteases are homologues of these other proteases in other parts of the fold.

183	SSSS	SSS	S	A						\$	A				TTTT	.T	T	tttt	.bt	T
184	KKKK	MMK	surface	A						A	AAAA	AAA	A		TTTT	TAT	T	TTTT	BbT	T
185	DDNA	RHD	SURFACE	A						A	AAAA	AAA	A		TTTT	TAT	T	TTTB	BbT	T
186	TDY	YYY	surface	A						A	AAAA	AAA	A		TTTT	TAT	T	BBtB	BbB	B
187	YYYY	YYY	inside	A						A	AAAA	AAA	A		BBTB	TAT		BBBB	BbB	B
188	QQQQ	QEQ	surface	A						A	AAAA	AAA	A		BBBB	TAT	B	BBBB	BAB	B
189	TTTT	KRM	surface	A						A	BBBB	BAA	B		BBBB	TAT	B	BBBB	BAB	B
190	FFFF	FFF	inside	A						A	BBBB	BBB	B		BBBB	TAT	B	BBBB	BAB	B
191	LLLL	LLL	inside	A						A	BBBB	BBB	B		TTBB	TTT	B	BBBB	BAB	B
192	TTTT	DKT	surface	A						A	BBBB	BBB	B		TTTT	TTT	T	BtBB	.AB	B
193	NNKD	QOK	SURFACE	A					p	A					TTTT	TTT	T	Bttt	TA.	T
194	HSYH	YR	SURFACE	A					p						.TTT	TTT	T	tttt	T..	T
195	NKKN	KKK	surface	A					p						tt..
196	PPEP	PPP	P	A					p						TTTT	TTT	T
197	QQQQ	QQQ	Q	a	B	\$	b	BBBB	BBB	B					TTTT	TTT	T	BBBB	BBB	B
198	CCCC	CCC	inside	a	B		b	BBBB	BBB	B					BBBB	TTT	B	BBBB	BBB	B
199	IIII	III	inside	a	B		b	BBBB	BBB	B					BBBB	TTT	B	BBBB	BBB	B
200	LILL	LLL	inside	a	B		b	BBBB	BBB	B					BBBB	TTT	B	BBBB	BBB	B
201	NNNN	NNN	N	a	B	\$	b	BB	BB						BBBB	...B		BBBB	BBB	B
202	AAAA	KKK	amphi	a	B		b									BBBB	...B	B
203	PPPP	PPP	P	A					p					

Figure 1 - Continued

REFERENCES

1. Benner, S. A., and Gerloff, D. (1991) *Adv. Enzyme Regulat.* 31, 121-181.
2. Benner, S. A., Cohen, M. A., and Gerloff, D. L. (1993) *J. Mol. Biol.* 229, 295-305.
3. Gerloff, D. L., Jenny, T. F., Knecht, L. J., Gonnet, G. H., and Benner, S. A. (1993) *FEBS Lett.*, 318, 118-124.
4. Benner, S. A. (1992) U.S. Patent Application, March 25, 1992.
5. Benner, S. A. (1992) *Curr. Opin. Struct. Biol.* 2, 402-412.
6. Knighton, D. R., Zheng, J., Ten Eyck, L., Ashford, F. V. A., Xuong, N. H., Taylor, S. S., and Sowadski, J. M. (1991). *Science* 253, 407-414.
7. Thornton, J. M., Flores, T. P., Jones, D. T., and Swindells, M. B. (1991). *Nature* 354, 105-106.
8. Lesk, A. M., and Boswell, D. R. (1992) *BioEssays* 14, 407-410.
9. Rost, B. Schneider, R., and Sander, C. (1993) *Trends Biochem. Sci.* 18, 120-123.
10. Robson, B., and Garnier, J. (1993) *Nature* 361, 506 (1993).
11. Benner, S. A. (1989) *Adv. Enzyme Regulat.* 28, 219-236.
12. Rost, B., and Sander, C. (1992) *Nature* 360, 540.
13. Rost, B., and Sander, C. (1993) *J. Mol. Biol.*, in press.
14. Chou, P. Y., and Fasman, G. D. (1978) *Adv. Enzymol.* 47, 45-148.
15. Garnier, J., Osguthorpe, D. J., and Robson, B. (1978). *J. Mol. Biol.* 120, 97-120.
16. Takeya, H., Arakawa, M., Miyata, T., Iwanaga, S., and Omori-Satoh, T. (1989) *J. Biochem. (Tokyo)* 106, 151-157.
17. Dayhoff, M. O., Schwartz, R. M., Orcutt, B. C., in "Atlas of Protein Sequence and Structure", M.O. Dayhoff, Ed., (National Biomedical Research Foundation, Washington, D.C., 1978) vol. 5, suppl. 3, p. 345 (1978).
18. Crawford, I.P., Niermann, T., and Kirschner, K. (1987) *Proteins* 2, 118-129.
19. Chothia, C., and Lesk, A. (1986) *EMBO J.* 5, 823-826.
20. Colman, P. M., Jansonius, J. N., and Matthews, B. W. (1972) *J. Mol. Biol.* 70, 701-724 (1972).
21. Bode, W., Gomis-Rueth, F. X., Huber, R., Zwilling, R., and Stoecker, W. (1992) *Nature* 358, 164-167 (1992).